

Copyright © 2019 by Academic Publishing House Researcher s.r.o.



Published in the Slovak Republic  
European Journal of Medicine  
Has been issued since 2013.  
E-ISSN: 2310-3434  
2019, 7(1): 45-56

DOI: 10.13187/ejm.2019.1.45  
[www.ejournal5.com](http://www.ejournal5.com)



## Effects of Catholyte Water on the Development of Experimental *Graffi* Tumor on Hamsters

Reneta Toshkova <sup>a</sup>, Elissaveta Zvetkova <sup>a</sup>, Ignat Ignatov <sup>b,\*</sup>, Georgi Gluhchev <sup>a</sup>

<sup>a</sup> Bulgarian Academy of Sciences (BAS), Bulgaria

<sup>b</sup> Scientific Research Center of Medical Biophysics (SRCMB), Bulgaria

### Abstract

The paper describes the results of a pilot study aimed at the investigation of the influence of catholyte (electrolyzed alkaline water) on the development of tumors. In the experiments solid *Graffi* tumor was transplanted subcutaneously in the back of the experimental group of *Golden Syrian* hamsters. Tumor appearance and growth were registered every day. Blood parameters were measured on the 10<sup>th</sup> and 25<sup>th</sup> day after transplantation and blood smears were prepared. Hamsters treated with catholyte developed tumor with some delay compared to untreated (drinking tap water) ones. Also, the tumor growth was slow and the survival rate was increased. The analysis of blood parameters and cell morphology has shown significant differences in the value of some hematometric parameters and morphological changes of some blood cells. The obtained results suggest about the beneficial influence of catholyte and the possible use of it as a supporting non-invasive therapy of cancer diseases.

**Keywords:** Graffi tumor, catholyte, tumor growth, survival rate, mortality, blood parameters.

### 1. Introduction

Water is a natural and necessary medium for many biological molecules. Alterations in its composition and structure can produce stimulating or inhibitory effects on the processes in the living things. Influenced by different factors water can change its acidity pH, ORP (Oxidation Reduction Potential), and its physical structure. When electrochemical activation or electrolysis is applied the obtained catholyte receives pH more than 8.0 and negative ORP in millivolts which leads to increased antioxidant effect (Prilutsky, Bakhir, 1997). Due to this reason it could be expected that the catholyte would have protective and positive effect for oxidative stress-related diseases like diabetes and cancer.

Clinical examinations carried out by different scientists have demonstrated positive effect in case of diabetes type 2, telomere shortening in cancer cells and inhibition of their growth, suppression of side effects caused by the use of anticancer medications, favorable influence on the blood system (Hayashi, Kawamura, 2002; Shirahata, 2012; Aschbah, 2019; Gluhchev, Ivanov, 2014; Sanetaka et al., 2012; Ignatov, Gluhchev., 2019). Along with this it was proved that the activated water was not toxic for cells and tissues, and did not have mutagenic, cancerogenic, embryotoxic or immunotoxic effects (Sanetaka et al., 2012).

\* Corresponding author

E-mail addresses: [mbioph@dir.bg](mailto:mbioph@dir.bg) (I. Ignatov)

For the evaluation of the influence of some medicine or therapy on tumor malignancies different parameters are used based on measurements of tumor development, survival rate, mortality, blood cells changes and others. In the last decade, the main hematometric indices (biomarkers) have been evaluated as diagnostic tools and prognostic parameters in patients with malignancies – cancer and leukemia (Zvetkova, Fuchs, 2017; Hirahara et al., 2016; Azab et al., 2013).

Recent data from the scientific oncological literature evaluated that the NLR (neutrophil to lymphocyte ratio) is superior for predicting the long term survival of cancer patients (Jia et al., 2015; Dezayee, Al-Nimer, 2016; Qin et al., 2018). E.g., lower NLR ( $\leq 2.0$ ) is associated with good prognosis for breast cancer patients; a higher peripheral blood NLR ( $\geq 5$ ) was considered to indicate – significantly and independently, a poor prognosis for breast cancer patients, gastric cancer patients, etc. Simultaneously, the total WBC count, neutrophil and lymphocyte counts alone could be also statistically significant predictors of 5-year cancer patients' mortality (Azab et al., 2013).

At present, the data about the effects of electrolyzed alkaline water (catholyte) on tumor growth and haematological parameters in experimental tumor-bearing animals are absent.

Recently the authors carried out a research about the influence of electromagnetic fields and infrared thermal fields on *Graffi* tumor (Toshkova et al., 2019).

The aim of this study is to examine the biological effects of catholyte on the tumor growth parameters, hematometric biomarkers (including main WBC count; granulocyte and lymphocyte count; lymphocyte percent, GLR (granulocyte to lymphocyte ratio), and blood cell morphology in hamsters with experimental *Graffi* myeloid tumor.

## 2. Materials and methods

*Experimental animals.* In the trials hamsters, breed “Golden Syrian”, aged 2-4 months, male and female, with weight around 100 g, grown in individual plastic cages with free access to food and water were used.

*Experimental tumor.* The experimental *Graffi* solid tumor is maintained on a monthly basis *in vivo* in hamsters from the research team at IEMPAM-BAS (Toshkova, 1995) via subcutaneous (**s.c.**) transplantation of live tumor cells ( $1-2 \cdot 10^6$ ) in the area of the back. Between days 7 and 15 in the spot of injection appear tumors, which grow progressively, and the hamsters die approximately 30-35 days after the injection. In such a tumor model it is observed 100 % appearance (transplantability) of tumor and 100 % mortality rate. Spontaneous regression, i.e. spontaneous shrinking and disappearance of the tumor is not observed.

*Catholyte water.* During the experiment catholyte water was produced every day using the Actvator-2 device, developed in the Institute of Information and Communication Technologies at the Bulgarian Academy of Sciences. In this way acidity pH of the water was kept between 9.0 and 9.5 and ORP  $< 0$ .

**2.1. Experimental design.** All the animals were divided into 4 groups as follows.

Gr.1 The hamsters from this group started drinking catholyte water 10 days before the injection with  $5 \times 10^4$  *Graffi* tumor cells per hamster in the back area, and continued drinking it until the end of the experiment.

Gr.2. This group was used as a control. The hamsters from it were **s.c.** injected with the same amount of tumor cells on the 10<sup>th</sup> day of the experiment as the hamsters from Gr.1, and were receiving tap water all the time.

Gr.3 consisted of healthy hamsters drinking catholyte water during the experiment.

Gr.4 consisted of healthy hamsters drinking tap water all the time.

The first two groups have to reveal the influence of the catholyte water on the appearance and growth of transplanted tumor compared to the tap water, as well as for evaluation of haematological parameters and peripheral blood cell morphology.

The last two groups were used as control for hematological research.

## 2.2. Measured parameters

The following parameters of tumor development are determined:

- tumor transplantability success (% of hamsters with tumor to the total number of injected ones),
- tumor size (the average diameter of tumor in mm measured with a caliper),
- survival and average survival (calculated for the respective group in days),
- lethality (% of dead animals in the group).

Animals from each group were sacrificed preserving the ethical aspects of the European convention for protection of vertebrate animals, used for experimental and other scientific purposes (OJ L 222), and approved by the National Veterinary Medical Office in Bulgaria, and blood samples have been prepared at different time periods: on days 10 and 25 after *Graffi* tumor implantation. Hematological/hematometric parameters and indices as shown in Fig.1 were measured on the automated hematological analyzer BC-2800 Vet (Mindray, China).

**2.3. The WBC/LR (White blood cells to Lymphocyte ratio and NLR (Neutrophil to lymphocyte ratio) were calculated.**

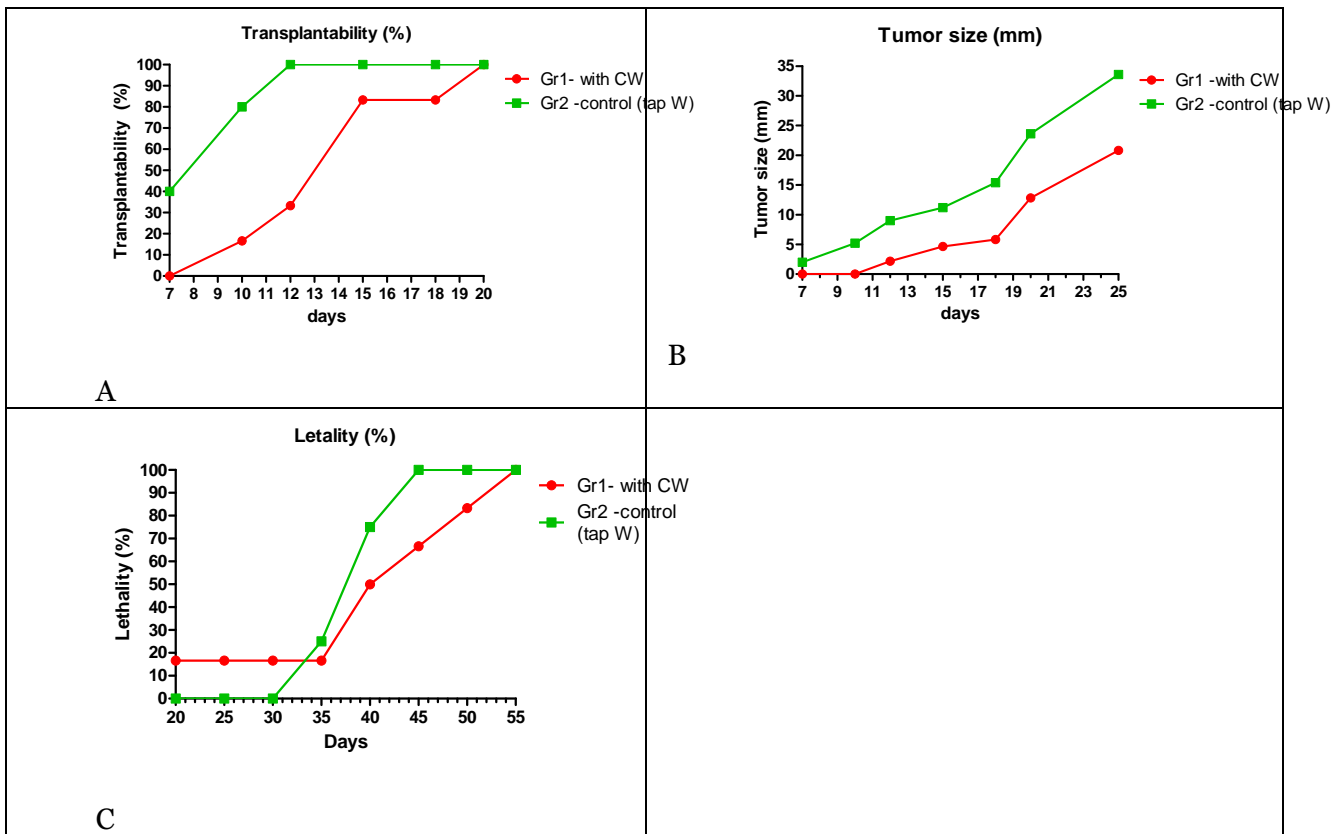
**2.4. Statistical analysis**

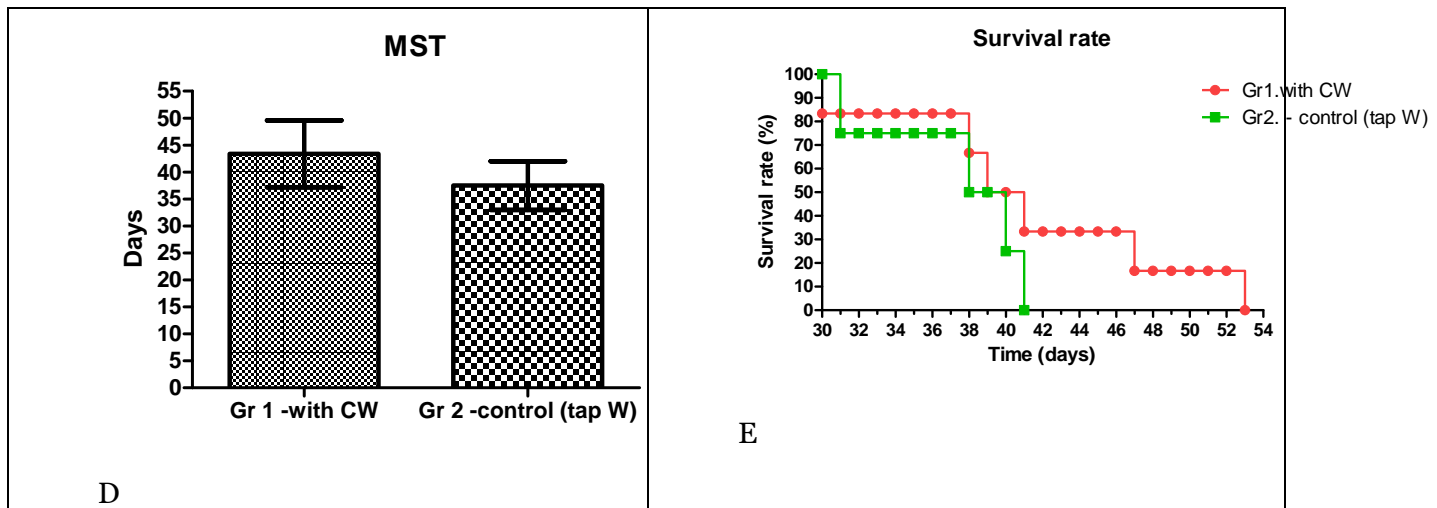
For the evaluation of the significance of the differences between the average values of a specific parameter *t*-test was used at levels of significance  $\alpha = 0.10$  and  $\alpha = 0.05$ , respectively.

**3. Results and discussion**

**3.1. Parameters of tumor growth for Gr.1 and Gr.2.**

Catholyte water was used as experimental therapy during two months. Animals have been examined every day until tumor detection and 2 times per week until 30 days after tumor transplantation. The tumor growth parameters have been registered regularly. The differences between the two groups are shown on (Figure 1).





**Fig. 1.** Biometric parameters of tumor growth for the hamsters from Gr.1 and Gr.2: A – transplatability in %; B – tumor size in mm; C – lethality in %; D – average survival time (AST) in days; E – Survival rate

As can be seen from the graph on (Figure 1A) the tumor transplantation was delayed for the hamsters taking catholyte. While all the hamsters from Gr.2 developed tumor on the 12<sup>th</sup> day this happened only for 1/3 of hamsters from Gr.1. In the hamsters receiving catholyte, tumors were detected at 100 % on day 20.

Similar effect is observed for the tumor size (Figure 1B). Until the 10<sup>th</sup> day subcutaneous firmness in the hamsters drinking catholyte was not established. This group showed an inhibition in tumor growth rate as compared to control throughout the study period.

The lethality for the hamsters from Gr.2 (Figure 1C) is increased after 35 days, compared with the hamsters from Gr.1. The control group (Gr.2) had a 100 % mortality on day 41, while the Gr.1 - on the 53<sup>rd</sup> day of the study.

The evaluated average survival for Gr.1 is  $43.4 \pm 6.9$  days, while it is  $37.5 \pm 4.5$  days for Gr.2. Same conclusion could be taken from (Figure 1E).

These data suggest that the catholyte water slows down tumor development and as a result increases survival rate.

The illustrations on (Figure 2) give visual impression for the tumor development in Gr. 1 and Gr. 2.



**Fig. 2.** Images of hamsters from Gr.1 (upper line) and Gr.2 (bottom line) taken on the 25<sup>th</sup> day after the tumor transplantation

### 3.2. Hematological parameters

Hamsters from the trial and control groups were euthanized after the application of deep anaesthesia on the 10<sup>th</sup> and 25<sup>th</sup> day after tumor transplanting. The obtained blood was used to report haematological parameters, for serum and preparation of blood smears, as well.

The evaluated parameters for all groups are displayed in (Table 1).

**Table 1.** Blood parameters of 10<sup>th</sup>/25<sup>th</sup> day of study

Parameters	Units	Tumor (Catholyte)	Tumor (Tap water)	Healthy (Catholyte)	Healthy (Tap water)
WBC (Leukocytes)	x10 <sup>9</sup> /L	3.8/ 47.9	5.7/2.9	6.8/5.8	2.4/2.1
Lymph	x10 <sup>9</sup> /L	2.2/38.1	0.7/0.5	5.6/2.1	1.4/1.2
Mon	x10 <sup>9</sup> /L	0.2/2.3	0.5/0.2	0.2/0.3	0.1/0.1
Gran	x10 <sup>9</sup> /L	1.4/ 7.5	4.5/2.2	1.0/3.4	0.9/0.8
Lymph %	%	58.3/79.5	12.8/16.5	81.4/35.5	58.8/56.3
Mon%	%	6.0/4.9	8.5/7.3	3.3/5.2	5.7/5.1
Gran%	%	35.7/ 15.6	78.7/76.2	15.3/59.3	35.5/38.6
RBC (Erythrocytes)	x10 <sup>12</sup> /L	3.05/2.95	4.77/5.58	4.7/7.98	4.25/5.36
HGB (Haemoglobin)	g/ L	67/83	80/104	85/137	89/ 92
HCT (Hematocrit)	L/L	0.165/0.204	0.231/0.325	0.267/0.445	0.226/0.304
MCV (Mean red blood cell volume)	fl	54.2/69.2	48.5/58.3	57.0/55.8	53.4/56.9
MCH (Average HGB content in erythr)	pg	21.9/28.1	16.7/18.6	18.0/17.1	20.9/17.1
MCHC(mean conc of Hb)	g/ L	406/406	346/320	318/307	393/302
RDW	%	12.6/22.9	14.1/14.0	15/11.7	12.8/16.1
PLT(Platelets)	x10 <sup>9</sup> /L	132/491	883/537	250/488	306/456
MPV(mean volume of platelets)	f/L	6.1/7.2	4.9/5.6	5.4/4.5	5.1/5.7
PDW	%	19.8/19.8	17.9/18.3	18.9/17.0	17.7/19.3
PCT	%	0.080/ 0.353	0.432/300	0.135/0.219	0.156/0.259



The development of experimental Graffi myeloid tumor in hamsters influenced diversely the two main WBC subpopulations – neutrophil granulocytes and lymphocytes (column 4). Significantly elevated WBC, granulocyte count and percent, as well as significant reduction of lymphocyte count and percent were observed in the Graffi myeloid tumor-bearing hamsters (Gr. 2, column 4) and (Figure 3). These effects are well expressed on the 10-th day, and are profound on the 25-th day. The treatment of tumor bearing animals with catholyte as drinking water improved the values of same parameters during the investigation (Gr.1, column 3) and (Figure 3).

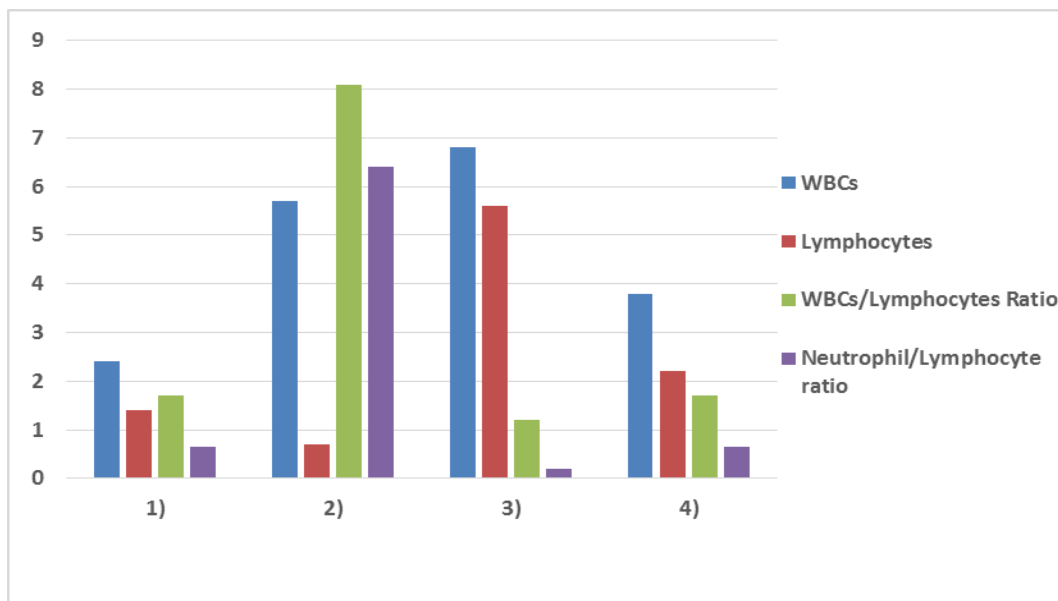
Results from the comparison of the blood parameters for which significant difference between Gr.1 and Gr.2 was obtained are shown in (Table 2).

**Table 2.** Significant difference between blood parameters of Gr.1 and Gr.2

Parameter	Lymp%	Mon%	Gran%	RBC	MCHC	PDW
$\alpha$	0.05	0.10	0.05	0.05	0.05	0.05

Based on haematological values the WBCs/LR (White blood cells to Lymphocyte ratio) and NLR (Neutrophil to Lymphocyte ratio) hematometric indices were calculated. Both WBCs/LR and NLR indices are strongly elevated in tumor hamsters taking tap water and highly reduced in hamsters taking catholyte water. The values are similar to healthy hamsters (Figure 3).

Differences in some of the hematological parameters (WBCs, Ly) and WBCs/LR, and NLR hematometric index for groups are shown in (Figure 3).



**Fig. 3.** WBCs (leukocytes) and Lymphocytes total count ( $\times 10^9 / L$ ), and WBCs/Lymphocytes Ratio (at the 10<sup>th</sup> day); 1) Controls; 2) Untreated tumor-bearing hamsters; 3) Healthy hamsters drinking catholyte; 4) Tumor-bearing hamsters drinking catholyte

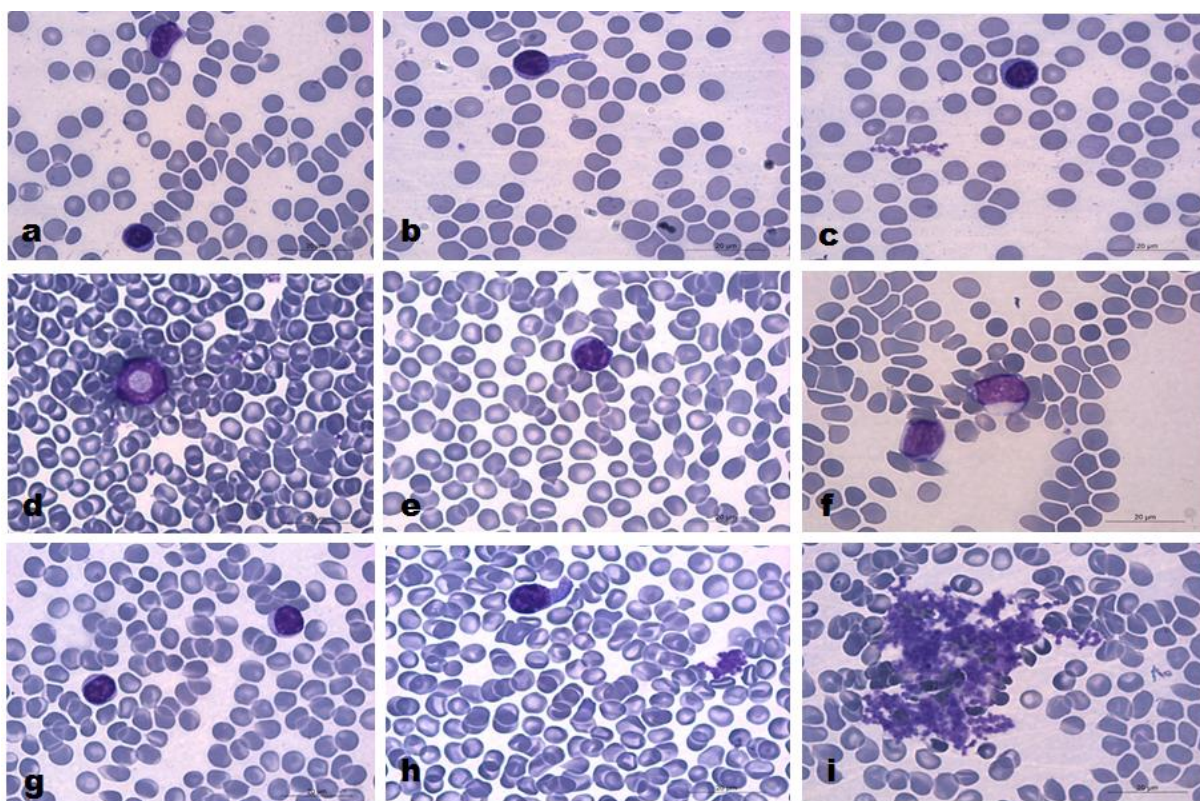
Significantly elevated WBC count and total granulocyte/neutrophil ratio were obtained in the untreated Graffi myeloid tumor-bearing hamsters (Gr. 2).

Simultaneously, a significant reduction of the lymphocyte number was evaluated in the same animals.

Additionally, we obtained that catholyte water influenced (elevated) some main PLT-hematometric values in both – control and experimental animals (Table 1, PLT, MPV and PDW). The hematometrical results obtained were confirmed by our cytological studies on PLTs in the peripheral blood smears of hamsters where one could see clusters of activated thrombocytes – more pronounced in the blood of tumor-bearing animals (Fig. 4 -i).

### 3.3. Cytological study

Images from blood smears are shown on (Figure 4).



**Fig. 4.** Images of blood smears as follows: upper line - healthy hamster treated with catholyte, middle line – tumor-bearing hamster (control) drinking tap water, and bottom line – tumor – bearing hamster treated with catholyte. (May Gruenwald Giemza staining. Objective X 100).

In the peripheral blood of healthy hamster, taking electrolyzed alkaline water (catholyte) activated lymphocytes with a large cytoplasmic pseudopode (ptotrusion), monocytes and platelets were observed (Figure 4 – a, b, c) upper line.

Atypical myeloid cells and blast-like cells from the peripheral blood of Graffi myeloid tumor-bearing hamsters at the day 10<sup>th</sup> from tumor inoculation were observed. One could see atypical immature granulocyte with peripherally localized ring-shaped nucleus and eosinophile granules – in the central part of cytoplasm (Figure 4 – d, e, f) middle line.

In Graffi tumor bearing hamsters, treated with catholyte atypical activated lymphocytes and cluster of PLTs in the peripheral blood smear were noticed. Lymphocytes from these hamsters, although atypical, show characteristic signs of activation as in healthy ones (Figure 4 – g, h, i) bottom line.

### 4. Conclusion

The influence of catholyte water on the development of *Graffi* tumor implanted in hamsters was assessed. Some delay in tumor growth and increased survival rate were observed. Significant differences in some of the blood parameters were noticed.

We obtained activated (small and medium-size) lymphocytes in the peripheral blood smears of healthy hamsters – treated with catholyte, instead of tap-water (Figure 4 – a, b, c). The same biological phenomenon was also evaluated partially in the peripheral blood of tumor-bearing animals, under the influence of catholyte (Figure 4-g, h). But in comparison to the activated immunocytes in healthy hamsters, the tumor-infiltration cells (TILs) in the tumor-bearing animals are soon atypical and insufficiently activated (or deactivated – in the preapoptotic or apoptotic states).

The appearance of the so-called blast-like cells (Figure 4 – f) has been related to dissemination of the neoplastic disease and could be earlier obtained in the peripheral blood

smears of untreated tumor-bearing animals (Zvetkova et al., 2006, 2007). The results correlated also with changes in WBCs/LR and NLR hematometric indices obtained in the two experimental groups of treated and untreated tumor-bearing hamsters.

The elevated thrombocytes count could be unfavorable predictor in cancer patients, having in view high risk of thrombogenesis and embolism. On the other hand, the catholyte water could be useful in cases with thrombocytopenia, but not in thrombocytoses, when application of electrolyzed water would be not recommended.

This study also further strengthens the role of WBC-hematometric indices in diagnosis and prognosis of cancer.

Catholyte water (investigated *in vivo* - in our experimental model of Graffi tumor-bearing hamsters), could improve TILs cellular immunity (immunomodulating, immunostimulating influence).

The first conclusion is that the developing experimental Graffi myloid tumor in hamsters influenced diversely the two main WBC subpopulations (predominantly neutrophils) and lymphocytes. These diverse effects – well expressed on the 10<sup>th</sup> day, are profound on the 25<sup>th</sup> day.

Our experimental results suggest that in this model the treatment of tumor-bearing animals with catholyte, as drinking water, improves the hematometric indices to the normal values.

Thus, our second conclusion is that the catholyte water – employed instead of tap water in our experimental model with tumor-bearing hamsters, has a positive impact on the main hematometric indices e.g. WBCs/LR and NLR - neutrophil to lymphocyte ratio on day 10 (Table 1) was: for WBCs/LR- 1.71 for a healthy hamster, 1.72 for a tumor-bearing, treated with catholyte and 8.14 for a tumor-bearing, untreated animal, and for NRL - 0.64 for a healthy hamster, 0.64 for a tumor-bearing, treated with catholyte and 6.42 for a tumor-bearing, untreated animal, respectively.

All these points at a favorable influence of catholyte on the hematopoiesis both in case of tumor-bearing animals, and healthy ones.

The obtained results lead to the general conclusion that catholyte could be used as a supporting non-invasive therapy to other cancer therapies as radiotherapy and chemotherapy. However, our pioneer study in this field needs further experimental and clinical confirmation.

## 5. Acknowledgements

The authors express thankfulness to Lieselotte Eder, Heide Trautwein, Gergana Genova, Paul N. Kleindienst, Borislav Kaleychev, Ivan Garabito for their support for the project.

## References

- Abdullah et al., 2012 – Abdullah, A.M., Abdelsalam, E., Abdullah, B., Khaled, A. (2012). Antioxidant Effects of Zamzam Water in Normal Rats and Those Under Induced-oxidant Stress. *Journal of Medicinal Plants Research*, 6(42), 5507-5512.
- Adelman et al., 1988 – Adelman, R., Saul, R., Ames, B. (1988). Oxidative Damage to DNA: Relation to Species Metabolic Rate and Life Span. *PNAS*, 85(8), 2706-2708.
- Aschbach, 2019 – Aschbah, D. (2019). Solutions of cations and anions with changeable ORP – new healing methods. Lecture at the seminar of Activated water at IICT-Bulgarian Academy of Sciences.
- Atanasov et al., 2014 – Atanasov, A., Karadzhov, S., Ivanova, E., Mosin, O.V., Ignatov I. (2014). Study of the Effects of Electrochemical Aqueous Sodium Chloride Solution (Anolite) on the Virus of Classical Swine Fever Virus. *Mathematical Models of Anolyte and Catolyte as Types of Water. Journal of Medicine, Physiology and Biophysics*, 4, 1-26.
- Azab et al., 2013 – Azab B. et al. (2013). Pretreatment Neurophil/lymphocyte Ratio is Superior to Platelet/lymphocyte Ratio as a Predictor of Long-term Mortality in Breast Cancer Patients. *Med. Oncol.*, 30, 2013, 432.
- Bakhrir, 1999 – Bakhrir, V.M. (1999). Theoretical Aspects of Electrochemical Activation. *2<sup>nd</sup> Int. Conf. Electrochemical Activation in Medicine, Agriculture and Industry*.
- Burger et al., 2012 – Burger, O., Baudish, A., Vaupel, J.W. (2012). Human Mortality Improvement in Evolutionary Context. *PNAS*, 109(44), 18210-18214.
- Chatzissavidou, Elke, 2001 – Chatzissavidou, N., Elke, L. (Eds) (2001). *Animal Cell Technology: From Target to Market*. Dordrecht. *Kluwer Academic Publishers*, 220-223.



Dezayee, Al-Nimer, 2016 – Dezayee, Z.M.I., Al-Nimer, M.S.M. (2016). The clinical importance of measurement of hematological indices in the breast cancer survival. *World J Oncol*, 7(1), 1-4.

Drossinakis et al., 2019 – Drossinakis, Ch., Toshkova, R., Zvetkova, E, Ignatov, I., Gluhchev, G. (2019). Methods of Research in Vivo Research of Therapeutical Effect in Hamsters with Experimental Myeloid Tumor of Graffi. 8th World Congress on Immunology, Pulsus, London, 3: 21.

Gluhchev et al., 2015 – Gluhchev, G., Ignatov, I., Karadzhov, S., Miloshev, G., Ivanov, N., Mosin, O.V. (2015). Virucidal and Bactericidal Effect of Anolyte and Catholyte Types of Water of Classical Swine Fever Virus and Bacteria E. Coli DH5. *Water: Hygiene and Ecology*, 3 (1-2): 5-23.

Gluhchev et al., 2015a – Gluhchev, G., Ignatov, I., Karadzhov, S., Miloshev, G., Ivanov, N., Mosin, O.V. (2015). Electrochemically Activated Water: Biophysical and Biological Effects of Anolyte and Catholyte Types of Water. *European Journal of Molecular Biotechnology*, 7(1): 12-26.

Gluhchev et al., 2015b – Gluhchev, G., Ignatov, I., Karadzhov, S., Miloshev, G., Ivanov, I., Mosin, O.V. (2015). Studying of Virucidal and Biocidal Effects of Electrochemically Activated Anolyte and Catholyte Types of Water on Classical Swine Fever Virus (CSF) and Bacterium E. coli DH5. *Journal of Medicine, Physiology and Biophysics*, 13: 1-17.

Gluhchev et al., 2015c – Gluhchev, G., Ignatov, I., Karadzhov, S., Miloshev, G., Ivanov, N., Mosin, O.V. (2015). Electrochemically Activated Water. Biophysical and Biological Effects of Anolyte and Catholyte as Types of Water. *Journal of Medicine, Physiology and Biophysics*, 10: 1-17.

Gluhchev et al., 2015d – Gluhchev, G., Ignatov, I., Karadzhov, S., Miloshev, G., Ivanov, N., Mosin, O.V. (2015). Studying the Antimicrobial and Antiviral Effects of Electrochemically Activated NaCl Solutions of Anolyte and Catholyte on a Strain of E. Coli DH5 and Classical Swine Fever (CSF) Virus. *European Journal of Medicine*, 9 (3): 124-138.

Gluhchev et al., 2015e – Gluhchev, G., Ignatov, I., Karadzhov, S., Miloshev, G., Ivanov, N., Mosin, O.V. (2015). Biocidal Effects of Electrochemically Activated Water.. *Journal of Health, Medicine and Nursing*, 11: 67-83.

Gluhchev et al., 2015f – Gluhchev, G., Ignatov, I., Karadzhov, S., Miloshev, G., Ivanov, N., Mosin, O.V. (2015). Antimicrobial and Antiviral Effects of Electrolytic Water Sodium Chloride (Anolyte/Catholyte) on a Strain of E.Coli DH5 and the Classical Swine Fever Virus (CSF). *Questions of Medical, Biological and Pharmaceutical Chemistry*, 3: 21-31.

Gluhchev, Ivanov, 2014 – Gluhchev, G., Ivanov, N. (2014). Electrochemically Activated Water. *J. of Ecological Engineering and Environment Protection*, 2: 68-73

Hirahara et al., 2016 – Hirahara N. et al. (2016). Prognostic Values of Hematological Parameters in Patients with Esophageal Squamous Carcinoma. *Int. J. Clin. Oncology*, 21(5), 909-919.

Ignatov et al., 2014 – Ignatov, I., Karadzhov, S., Atanasov, A., Ivanova, E., Mosin, O.V. (2014). Electrochemical Aqueous Sodium Chloride Solution (Anolyte and Catholyte) as Types of Water. Mathematical Models. Study of Effects of Anolyte on the Virus of Classical Swine Fever Virus. *Journal of Health, Medicine and Nursing*, 8: 1-28.

Ignatov et al., 2015 – Ignatov, I., Gluhchev, G., Karadzhov, S., Miloshev, G., Ivanov, N., Mosin, O.V. (2015). Preparation of Electrochemically Activated Water Solutions (Catholyte/Anolyte) and Studying Their Physical-Chemical Properties. *Journal of Medicine, Physiology and Biophysics*, 11: 1-21.

Ignatov et al., 2015a – Ignatov, I., Gluhchev, G., Karadzhov, S., Miloshev, G., Ivanov, N., Mosin, O.V. (2015). Preparation of Electrochemically Activated Water Solutions (Catholyte/Anolyte) and Studying of their Physical-Chemical Properties. *Journal of Medicine, Physiology and Biophysics*, 13: 18-38.

Ignatov et al., 2015b – Ignatov, I., Gluhchev, G., Karadzhov, S., Miloshev, G., Ivanov, N., Mosin, O.V. (2015). Preparation of Electrochemically Activated Water Solutions (Catholyte/Anolyte) and Studying of their Physical-Chemical Properties. *Journal of Health, Medicine and Nursing*, 13: 64-78.

Ignatov et al., 2015d – Ignatov, I., Mosin, O. V. , Gluhchev, G., Karadzhov, S., Miloshev, G., Ivanov, N. (2015). Studying Electrochemically Activated NaCl Solutions of Anolyte and Catholyte by Methods of Non-Equilibrium Energy Spectrum (NES) and Differential Non-Equilibrium Energy Spectrum (DNES). *Journal of Medicine, Physiology and Biophysics*, 14: 6-18.

[Ignatov et al., 2015e](#) – Ignatov, I., Gluhchev, G., Karadzhov, S., Ivanov, N., Mosin, O.V. (2015). Preparation of Electrochemically Activated Water Solutions (Catholyte/Anolyte) and Studying Their Physical-Chemical Properties. *Journal of Medicine, Physiology and Biophysics*, 16: 1-14.

[Ignatov et al., 2015f](#) – Ignatov, I., Mosin, O.V., Gluhchev, G., Karadzhov, S., Miloshev, G., Ivanov, N. (2015). The Evaluation of Mathematical Model of Interaction of Electrochemically Activated Water Solutions (Anolyte and Catholyte) with Water. *European Reviews of Chemical Research*, 2 (4): 72-86.

[Ignatov et al., 2015g](#) – Ignatov, I., Mosin, O.V., Gluhchev, G., Karadzhov, S., Miloshev, G., Ivanov, N. (2015). The Evaluation of Mathematical Model of Interaction of Electrochemically Activated Water Solutions (Anolyte and Catholyte) with Water. *European Reviews of Chemical Research*, 2 (4): 72-86.

[Ignatov et al., 2016](#) – Ignatov, I., Mosin, O.V., Gluhchev, G., Karadzhov, S., Miloshev, G., Ivanov, N. (2016). Studying Electrochemically Activated NaCl Solutions of Anolyte and Catholyte by Methods of Non-Equilibrium Energy Spectrum (NES) and Differential Non-Equilibrium Energy Spectrum (DNES). *Journal of Medicine, Physiology and Biophysics*, 20: 13-23.

[Ignatov et al., 2016a](#) – Ignatov, I., Mosin, O.V., Kirov, P. (2016). Mathematical Model of Kangen Water® Biophysical and Biochemical Effects of Catholyte. *Advances in Physics Theories and Applications*, 51: 33-55.

[Ignatov et al., 2018](#) – Ignatov, I., Karadzhov, S., Gluhchev, G., Yakimov, I. (2018). Electromagnetically Activated Water – Properties and Effects. *Bulgarian Journal of Public Health*, 10(4): 63-69.

[Ignatov et al., 2019](#) – Ignatov, I., Toshkova, R., Gluhchev, G., Drossinakis, Ch. (2019). Results of Blood Serum from Cancer Treated Hamsters with Infrared Thermal Field and Electromagnetic Fields. *Journal of Health, Medicine and Nursing*, 58: 101-112.

[Ignatov, Gluhchev, 2019](#) – Ignatov, I., Gluhchev, G. (2019). Effects of Electrochemically Activated Water Catholyte and Anolyte on Human Health. *8th World Congress on Immunology, Pulsus, London*, 3: 12-13.

[Ignatov, Mosin, 2013](#) – Ignatov I., Mosin O.V. (2013). Possible Processes for Origin of Life and Living Matter with Modeling of Physiological Processes of Bacterium *Bacillus Subtilis* in Heavy Water as Model System. *Journal of Natural Sciences Research*, 3 (9), 65-76.

[Ignatov, Mosin, 2013](#) – Ignatov, I., Mosin, O.V. (2013). Structural Mathematical Models Describing Water Clusters. *Journal of Mathematical Theory and Modeling*, 3 (11): 72-87.

[Ignatov, Mosin, 2013a](#) – Ignatov, I., Mosin, O.V. (2013). Modeling of Possible Processes for Origin of Life and Living Matter in Hot Mineral and Seawater with Deuterium. *Journal of Environment and Earth Science*, 3(14): 103-118.

[Jia et al., 2015](#) – Jia W. et al. (2015). Neutrophil-to-lymphocyte Ratio is Superior to the Lymphocyte –to-Monocyte Ratio for Predicting the Long Term Survival of Triple Negative Breast Cancer Patients. *PlosOne*, 10(11).

[Kirkpatrick, 2009](#) – Kirkpatrick, R.D. (2009). The Mechanism of Antimicrobial Action of Electro-chemically Activated (ECA) Water and its Healthcare Applications. Doctoral Thesis, University of Pretoria.

[Kloss, 1988](#) – Kloss, A.I. (1988). Electron-radical Dissociation and Mechanism of Water Activation. *Trans. Acad. Sc. USSR*, 303: 1403-1406. [in Russian]

[Kokichi et al., 2004](#) – Kokichi H., Dongxu S., R., Lawrence, Y. Kamitani, Fernandes G. (2004). The Mechanism of the Enhanced Antioxidant Effects Against Superoxide Anion Radicals of Reduced Water Produced by Electrolysis. *Biophysical Chemistry*, 107: 71-82.

[Komatsu et al., 2001](#) – Komatsu, T., Kabayama, S., Hayashida, A et al. (2001). Suppressive Effect of Electrolyzed Reduced Water on the Growth of Cancer Cells and Microorganisms. In E. Lindner-Olsson.

[Kumar et al., 1999](#) – Kumar, S.V., Ezeike G.O., Hung, Y-C., Doyle M.P. (1999). Efficacy of Electrolyzed Oxidizing Water for Inactivating Escherhia coli O157:H7, Salmonela enteritidis, and Lusteria monocytogenes. *Applied and Environmental Microbiology*: 4276-4279.

[Kyu-Jae Lee et al., 2004](#) – Kyu-Jae, Lee, Seung-Kyu, Jae-Won, Gwang-Young et al. (2004). Anticancer Effect of Alkaline Water, Korea.

Lee et al., 2006 – Lee, M-Y., Kim.-K., Ryoo, K.-K et al. (2006). Electrolyzed-reduced water protects against oxidative damage to DNA, RNA and protein. *Applied Biochemistry and Biotechnology*, 135: 133-144.

Lv et al., 2011 – Lv. J., Wang, W., Krafft, T., Li, Y., Zhang, F., Yuan, F. (2011). Effects of Several Environmental Factors on Longevity and Health of the Human Population of Zhongxiang, Hubei, China. *Biol. Trace Elem. Res.*, 143 (2): 702-716.

Mariani et al., 2006 – Mariani, E. et al. (2006). Antioxidant Enzyme Activities in Healthy Old Subjects: Influence of Age, Gender and Zinc Status: Results from the Zincage Project. *Biogerontology*, 7(5-6): 391-398.

Mehandjiev et al., 2017 – Mehandjiev, D., Ignatov, I., Karadzhov, I., Gluhchev, G., Atanasov, A. (2017). On the Mechanism of Water Electrolysis. *Journal of Medicine, Physiology and Biophysics*, 31: 23-26.

Mehandjiev et al., 2017 – Mehandjiev, D., Ignatov, I., Karadzhov, S., Gluhchev, G., Atanasov, A. (2017) Processes in Catholyte and Anolyte as Result of Water Electrolysis. *European Journal of Molecular Biotechnology*, 5 (1): 23-29.

Miroshnikov, 2002 – Miroshnikov A.I. (2002). Stimulation and Inhibition of Escherichia coli Cell Growth During Cultivation in the Catholyte and Anolyte of Culture Medium. *Biofizika*, 47(2): 304-308. [in Russian]

Petrushenko, Lobyshev, 2004 – Petrushenko, I., Lobyshev, V.I. (2004). Physico-chemical Properties of Aqueous Solutions, Prepared in a Membrane Electrialyzer. *Biofizika*, 49(1): 22-31. [in Russian]

Popova et al., 2016 – Popova, T., Petrova, T., Karadzhov, S. (2016). Investigation of Biocidal Effect of Electrochemically Activated Aqueous Sodium Chloride on Gram-negative Pathogenic Bacteria. *Int. J. Curr. Microbiol. App. Sci*, 5(1): 624-652.

Popova et al., 2016 – Popova, T., Petrova, T., Karadzhov, S. (2016). Investigation of the Action of the Anolyte after Different Storage Times in the Gram negative Bacteria. *Int. J. Curr. Microbiol. App. Sci*, 5(9): 530-539.

Popova et al., 2016 – Popova, T., Petrova, T., Karadzhov, S., Krustanova, G. (2016). Investigation of the Biocidal Effect of Electrochemically Activated Aqueous Sodium Chloride Solution on Staphylococcus Aureus. *Tradition and Modernity in Veterinary Medicine*, 1(1): 67-72.

Prilutsky, Bakhir, 1997 – Prilutsky, V.I., Bakhir V.M. (1997). Electrochemically Activated Water: Anomalous Properties, Mechanism of Biological Action. *All Russian Scientific Research and Experimental Institute of Medical Engineering (VNIIMT)*, 1: 124. [in Russian]

Qin Y-y et al., 2018 – Qin Y-y et al. (2018). Red Blood Cell Distribution Width, Mean Platelet Volume and Cancer Antigen-125 as Inflammatory Markers in Differential Diagnosis of Ovarian Cancer. *J Ovarian Res.*

Shirahata et al., 2012 – Shirahata, S., Hamasaki, T., Teruya, K. (2012). Advanced Research on the Health Benefit of Reduced Water. *Trends in Food Science & Technology*, 23: 124-131.

Shirahata, 2000 – Shirahata, S. (2000). Reduced Water for Prevention of Diseases. *Animal Cell Technology: Basic & Applied Aspects*: 25-30.

Suzuki et al., 2002 – Suzuki, T., Itakura, J., Watanabe, M., Ohta, M., Sato, Y., Yamata, Y. (2002). Inactivation of Staphylococcal Enterotoxin-A with an Electrolyzed Anodic Solution. *Journal of Agricultural and Food Chemistry*, 50: 230-234.

Tanaka et al., 1996 – Tanaka, H., Hirakata, Y., Kaku, M., Yoshida, R., Takemura, H., Mizukane, R. (1996). Antimicrobial Activity of Superoxidized Water. *Journal of Hospital Infection*, 34(1): 43-49.

Toshkova et al., 2019 – Toshkova, R., Ignatov, I., Zvetkova, E., Gluhchev, G. (2019). Bioinfluence with Infrared Thermal and Electromagnetic Fields as a Therapeutic Approach of Hamsters with Experimental Graffi Myeloid Tumor. *Journal of Natural Sciences Research*, 9(4): 1-11.

Toshkova et al., 2019 – Toshkova, R., Ignatov, I., Zvetkova, E., Gluhchev, G., Drossinakis, Ch. (2019). Beneficial Effects of Drossinakis Bio-influence (with Infrared Thermal and Electromagnetic Fields) on the Development of Experimental Graffi Myeloid Tumors in Hamsters. Hematological studies. *Journal of Medicine, Physiology and Biophysics*, 54: 13-17.

Toshkova, 1995 – Toshkova, R. (1995). Attempts for Immunomodulation in Hamsters with Transplanted Myeloid Tumor, Previously Induced by Graffi Virus. Bulgarian Academy of Sciences, PhD Dissertation, Sofia.

[Vassileva et al., 2019](#) – Vassileva, P., Voykova, D., Ignatov, I., Karadzhov, S., Gluhchev, G., Ivanov, N., Mehandjiev, D. (2019). Results from the Research of Water Catholyte with Nascent (Atomic) Hydrogen. *Journal of Medicine, Physiology and Biophysics*, 52: 7-11.

[Yahagi et al., 2000](#) – Yahagi, N., Kono, M., Kitahara, M., Ohmura, A., Sumita, O., Hashimoto, T. (2000) Effect of Electrolyzed Water on Wound Healing. *Artificial Organs*, 24(12): 984-987.

[Ye et al., 2004](#) – Ye, J., Teruya, K., Katakura, Y. et al. (2004). Suppression of Invasion of Cancer Cells and Angiogenesis by Electrolyzed Reduced Water. *World Congress on in Vitro Biology*.

[Ye et al., 2008](#) – Ye, J., Li, Y., Hamasaki, T., Komatsu, N. et al. (2008). Inhibitory Effect of Electrolyzed Reduced Water on Tumor Angiogenesis. *Biological and Pharmaceutical Bulletin*, 31: 19-26.

[Zinkevich, 2000](#) – Zinkevich, V., Beech, I.B., Tapper, R.C., Bogdarina, I. (2000). The Effect of Super-oxidized Water on Escherichia Coli. *Journal of Hospital Infection*, 46(2): 153-156.

[Zvetkova, 2006](#) – Zvetkova E. (2006). F.140. Quantitative Reduction in the RNP-Contents of Peripheral Blood Lymphocytes in Cancer Patients. Conference: 6<sup>th</sup> Annual Meeting of the Federation-of-Clinical-Immunology-Societies Location: San Francisco, CA Date: JUN 07-11, Clinical Immunology: 119: S100.

[Zvetkova, Fuchs, 2017](#) – Zvetkova, E., Fuchs, D. (2017). Medical Significance of Simultaneous Application of Red Blood Cell Distribution Width (RDW) and Neopterin as Diagnostic Biomarkers in Clinical Practice. *Pteridines*, 28(3-4): 133-140.